Optimum utilization of brewers' yeast (*Saccharomyces cerevisiae*) to replace local fish meal in juvenile Sea cucumber (*Holothuria scabra*) diets

R. Weerasingha^{1*}, M. S. M. Fahim¹, P.A.D. A. Kumara¹, D. R. Kumarasinghe² and C. B. Madagedara¹

¹National Aquatic Resources Research and Development Agency (NARA), Crow Island, Colombo 15, Sri Lanka

²Department of Animal Science, Faculty of Agriculture, University of Peradeniya, Sri Lanka

Abstract

The juvenile sea cucumber, *Holothuria scabra* was reared for 55 days in a re-circulatory tank system consisting of filtered seawater flowing at 2.5-3 L/min. One hundred and forty-four juveniles of an average weight of 0.88 ± 0.03 g were randomly distributed in 12 tanks of 250 L. Sea cucumber were fed four diets containing different dietary concentrations of brewer's yeast (BY), *Saccharomyces cerevisiae* (BY₀: 0%; BY₁₀: 10%; BY₃₀: 30%; BY₅₀: 50%) to replace the local fish meal (CM fish meal[®]). At the end of the experiment, the specific growth rate (SGR), weight gain (WG) and final body weight (FW) of juvenile sea cucumber fed with BY₁₀ was significantly higher than those of juveniles fed BY₀ and BY₅₀ (p<0.05) though, not significantly higher than those fed BY₃₀ (p>0.05). The optimum replacement level of CM fish meal[®] with brewers' yeast was 13.75% for juvenile *H. scabra* by polynomial regression analysis of WG.

Keywords: Brewer's yeast, *Saccharomyces cerevisiae*, local fishmeal, juvenile sea cucumber, *Holothuria scabra*

^{*}Corresponding author email: rochanaweerasingha@gmail.com

Introduction

Sea cucumber became a high-demanded commodity among all seafood varieties in the world after the new trade economy of China started in the 1980's (Hamel *et al.*, 2001). Many coastal nations lost their wild sea cucumber population due to overexploitation (Ajith Kumara *et al.*, 2013; Battaglene *et al.*, 1999; Hamel *et al.*, 2001; Purcell *et al.*, 2012). Hence, the larval production of *Holothuria scabra* increased gradually in commercial hatcheries and it cost a high market value. The species is naturally distributed in shallow coastal areas across the Indo-Pacific region. As the pioneer in the South Asian region, India started to produce *H. scabra* seeds while the wild stock was declining (James, 2004). The production of *H. scabra* juveniles in Sri Lankan hatcheries was initiated in 2011 (Kumara and Dissanayake, 2017). The processed sea cucumber production collected from wild and pen cultured in Sri Lanka ranged from 258 tons in 2005 to 248 tons in 2018, with several annual fluctuations (DOF, 2019). The highest value reported was 7.65 USD million in 2018 (DOF, 2019).

In general, dried and powdered seaweed are used as feeds in sea cucumber hatcheries. Seasonality issues in the availability of seaweed, drying issues, and nutrient imbalances occur in feeding seaweed for sea cucumber juveniles. The predominant protein source in aqua feeds is still fishmeal (FM). The fish meal provides adequate levels of amino acids for aquaculture species. The negative impacts, such as imbalances in amino acid profile, reduced palatability, and availability of anti-nutritional factors, have been reported in using plant protein sources as alternative sources (NRC, 2011). Therefore, higher levels of plant protein sources to replace the fish meal are also impractical. However, soybean meal replaced 40% of fishmeal in sea cucumber, *Apostichopus japonicus* juvenile diets without affecting growth (Fan *et al.*, 2010). In contrary, 60% FM replacement by soybean meal did not affect growth in sea cucumber, *Apostichopus japonicus* juveniles (Liao *et al.*, 2015). Other alternative fish meal replacers in sea cucumber diets such as silkworm caterpillar meal (Sun *et al.*, 2014) and alga *Spirulina* meal (*Tan et al.*, 2009) were suggested to be good fish meal replacers in sea cucumber, *Apostichopus japonicus japonicus* juveniles is place.

Saccharomyces species are used in the brewing industry to produce beer from wort (Bourdichon *et al.*, 2012). The fermented product in beer production called brewers'

yeast (BY) has been studied as a dietary supplement and growth promoter for hybrid striped bass, Morone chrysops× M. saxatilis (Li and Gatlin, 2003) and hybrid of African catfish, Clarias gariepinus (Essa et al., 2011), as an immune stimulant for hybrid striped bass (Li & Gatlin, 2004), and as an FM replacer for sea bass, Dicentrarchus labrax juveniles (Oliva-Teles & Gonçalves, 2001), goldfish, Carassius auratus (Gumus, Aydin, & Kanyilmaz, 2016) and pacu, Piaractus mesopotamicus (Ozório et al., 2010). The studies on the utilization of brewers' yeast products suited for aqua feeds have been expanded up to compounds contained in brewers' yeast, Saccharomyces cerevisiae. For instance, glucan derived from S. cerevisiae (bakers' yeast) was injected to channel catfish, Ictalurus punctatus and improved disease tolerance against Edwardsiella ictaluri (Chen & Ainsworth, 1992). However, partial substitution of dietary baker's yeast S. cerevisiae did not affect trout fish growth (Rumsey, Hughes, & Kinsella, 1990). Dietary addition of S. cerevisiae in rainbow trout, Oncorhynchus mykiss diets enhanced disease resistance against Aeromonas salmonicida (Siwicki, Anderson, & Rumsey, 1994) and improved growth and immunity with disease resistance against Yersinia ruckeri (Tukmechi et al., 2011).

Brewers' yeast grouped into the single-cell protein (SCP) category gives many advantages to fish feeds (Couttau & Lavens, 1989). Single-cell proteins like microalgae, yeast and bacteria are utilized in fish feeds considering their nutrients like proteins, pigments, B-vitamins, and carbohydrates (especially glucans) (Sanderson & Jolly, 1994;Tacon, 1995). B-glucan consisted in the yeast cell wall is an immunostimulant/ feed additive for aquaculture organisms (Sakai, 1999; Thanardkit *et al.*, 2002). Generally, microbial products or fractions like barley and brewers'yeast are utilized for the β -glucan extraction (NRC, 2011). Likewise, sea cucumber, *A. japonicus* juveniles fed diets containing dietary β -glucan showed an improved immune response and balance of intestinal microbiota (Yang *et al.*, 2015).

Therefore, we aimed to find the optimum supplement level of brewers' yeast, *S. cerevisiae* by replacing a local FM in juvenile sea cucumber (*H. scabra*) diet to maximize their growth.

Materials and Methods

Experimental site

The experiment was carried out in a re-circulatory tank system of 12 fiberglass tanks of 250 L at the hatchery of the regional research center, National Aquatic Resources Research and Development Agency (NARA), Kalpitiya, Sri Lanka.

Experimental feeds

The main ingredients, a local fish meal/ CM fish meal[®] (CMFM) procured from Cool man fish meal factory, Pesalei and brewers' yeast provided by Lion Brewery (Ceylon) PLC, Biyagama were used with all other ingredients that purchased from the retail market, Pettah. Soybean meal, meat and bone meal, maize, poonac, rice polish and brewers' yeast were ground separately by a hammer mill and sieved with a 0.5 mm sieve. Four diets (BY₀: 0% brewers' yeast (BY); BY₁₀: 10% CMFM replaced by BY; BY₃₀: 30% CMFM replaced by BY; BY₅₀: 50 % CMFM replaced by BY) were formulated and prepared practical diets (Table 2). Then, prepared feeds were stored at – 20 °C until the commencement of the feed trial.

	Crude	Moisture%	Ash%	Fiber%	Fat%
	Protein%				
CMFM	42.37	2.9	33.9	1.4	5.92
BY	39.3	6.0	6.5	4.3	0.7

 Table1. Proximate composition of dry brewers'yeast (Saccharomyces cerevisiae)

Experimental Design

One hundred and forty-four sea cucumber juveniles produced in SL Aquatech International (Pvt.) Ltd., Chilaw averaging 0.88 ± 0.04 g size were placed in 12 fiberglass tanks of 250 L capacity at a stocking density of 12 juveniles per tank. The four experimental diets were randomly assigned in triplicated tanks.

Feeding trial

Prior to the commencement of the trial *H.scabra* juveniles were acclimated to the tank system and feeds for a week. The culture system was operated with filtered seawater for 55 days of the feeding trial. The re-circulatory system was operated 2 hrs per day at a 2.5-3 L/ min flow rate and water was exchanged three times per week. Salinity, dissolved oxygen (DO) and ionized ammonia NH_{3+} were checked daily while the values were maintained 25-28ppt, 5.5-6 mg/ L and 0.07-0.10 mg/ L respectively. Feeding proceeded once a day at 4% of the body weight. Growth data were collected biweekly to adjust the feed amount.

Ingredient	$\mathbf{B}\mathbf{Y}_0$	BY10	BY ₃₀	BY ₅₀
$\mathrm{CMFM}^{\mathbb{R}^1}$	20	18	14	10
Brewers'yeast ²	0	2	7	10
Soybean meal ³	20	20	20	20
Meat and bone meal ³	5	5	5	5
Maize ³	15	15	15	15
Poonac ³	5	5	5	5
Rice polish ³	5	5	5	5
Wheat flour ³	26	26	25	26
Fish oil ³	0.5	0.5	0.5	0.5
Vitamin mineral premix ^{3,4}	3	3	3	3
DL-Methionine ³	0.3	0.3	0.3	0.3
L-lysine ³	0.2	0.2	0.2	0.2
Proximate composition (DM basis)				
Moisture	9.32±0.35	9.21±0.79	8.82±0.01	9.01±0.11
СР	29.89±1.01	29.67±0.97	29.94±0.25	29.12±0.24
Ash	16.53±0.51	14.57±0.29	13.52±0.16	11.92±0.02
Crude Fat	3.17±0.24	2.97±0.27	2.64±0.32	2.31±0.15
Crude Fiber	2.75±0.14	2.86±0.11	3.08±0.10	3.30±0.09
NFE	38.34±0.23	40.72±0.49	42.6±0.20	44.34±0.13

Table 2. Composition of experimental feeds and their proximate composition (means \pm SE)

¹Cool Man fish meal[®] (CMFM) was procured from the cool man fish meal factory, Thaleimannar road, Pesalei, Sri Lanka.

² Brewers' yeast (BY) was provided by Lion Brewery (Ceylon) PLC, Colombo road, Biyagama, Sri Lanka.

³All the other ingredients were purchased from the retail market, Pettah, Sri Lanka.
⁴ Contains (as mg/kg in diets): Vitamin A, 9000IU; Vit K, 2 mg; Vit E, 5 mg; Vit B1, 2 mg; Vit B2, 3.6 mg; Vit B6, 1 mg; Vit B12, 10 mg; Vit D3, 2000 IU; Chlorine chloride,150 mg; Mn,60 mg; Zn, 50 mg; Fe, 25 mg; Niacinamide, 16 mg; I,5.5 mg; Cu,5 mg;

Calcium pantothenate, 4 mg; Folic acid, 0.5 mg; CO, 0.1 mg.

Statistical Analysis

Growth performance data were subjected to one-way analysis of variance (ANOVA) tests at p<0.05 level of significance. If there is a significant difference, the treatments were compared by Tukey's test at p<0.05 using SAS 9.4 program.

Results and Discussion

During feeding trial, *H. scabra* juveniles accepted all the experimental feeds well. At the end of the experiment, a significantly higher specific growth rate (SGR), weight gain (WG) and final body weight (FW) showed by the juvenile sea cucumber fed BY_{10} followed by BY_{30} compared to juveniles fed BY_0 and BY_{50} (p<0.05). However, there were no significant differences in the growth performances of juveniles fed BY_{30} and BY_{10} (p>0.05) (Table 3).

For the last decades, the attention for brewers' yeast to be used in aquafeeds has increased. As it is a byproduct in the beer industry (Bourdichon *et al.*, 2012) which consisting of protein, vitamin B complex and β -glucan (Sanderson & Jolly, 1994;Tacon, 1995), brewers' yeast has been suggested utilizing in aquafeeds as a feed additive and fish meal replacer. In our experiment, the dietary addition of brewers' yeast improved the growth of *H. scabra* juveniles. So far, the utilization of dietary brewers' yeast, *S.cerevisiae* in sea cucumber *H. scabra* juvenile diets has not been reported. Benefits of using brewers' yeast in the diets of other cultured species like freshwater prawn

Macrobrachium rosenbergii post larvae (Prasad *et al.*,2013) and finfish like sea bass *Dicentrarchus labrax* juveniles (Oliva-Teles & Gonçalves, 2001), hybrid striped bass *Morone chrysops* \times *M. saxatilis* (Li & Gatlin, 2003),(Li & Gatlin, 2004), hybrid of African catfish *Clarias gariepinus* (Essa *et al.*,2011), pacu *Piaractus mesopotamicus* (Ozório *et al.*, 2010) and goldfish *Carassius auratus* (Gumus *et al.*, 2016) were reported. In the study, 10% dietary addition of brewers' yeast enhanced growth performance results; though not significantly better than 30% brewers' yeast. Ozório *et al.* (2010) stated that brewers' yeast could replace 50% FM in pacu *Piaractus mesopotamicus* diets without affecting growth.

Table 3. Growth performances of *H. scabra* juveniles fed four different feeds (means \pm SE) (n=3)¹

		Diets		
—	$\mathbf{B}\mathbf{Y}_0$	BY ₁₀	BY ₃₀	BY ₅₀
Initial weight (g)	0.86±0.00	0.86 ± 0.00	0.91±0.01	0.89 ± 0.00
Final weight (g)	1.63±0.15 ^b	2.39±0.12 ^a	1.97±0.09 ^{ab}	1.73 ± 0.15^{b}
WG (%) ²	$81.03{\pm}16.82^{\text{b}}$	165.61±12.88 ^a	119.08±9.71 ^{ab}	$92.30{\pm}17.14^{b}$
$SGR(\%day^{-1})^3$	1.06±0.17 ^b	1.77±0.09 ^a	1.42±0.08 ^{ab}	1.17±0.17 ^b
Survival (%) ⁴	58.33±16.67	52.78±2.78	63.89±7.35	55.56±10.02

¹Values in each column with different superscripts are significantly different (p<0.05).

²Weight gain = $\left[\frac{\text{final weight-initial weight}}{\text{initial weight}}\right] x100$ ³Specific growth rate (SGR) = $\frac{(\text{Ln})\text{final weight}-(\text{Ln})\text{initial weight})}{\text{days}} x 100$

 ${}^{4}Survival = \frac{No.of juveniles at the start-No.of juveniles at the end}{No.of juveniles at the start} x 100$

The optimum replacement level of CMFM with brewers' yeast for juvenile sea cucumber, *H. scabra* was 13.5% by the polynomial analysis of WG (Fig. 1).



Fig. 1. Optimum replacement level of CMFM by brewers' yeast on weight gain of sea cucumber, *Holothuria scabra* juveniles fed 55 days.

Moreover, in goldfish *Carassius auratus* diets, brewers' yeast was able to replace 35% FM without adverse effects on growth (Gumus *et al.*, 2016). Oliva-Teles & Gonçalves (2001) mentioned that brewers' yeast could replace 50% of FM in *Dicentrarchus labrax* juvenile diets with no effect on growth performances. Furthermore, 30% of dietary brewers' yeast in *Dicentrarchus labrax* diets enhanced protein utilization and feed efficiency. Likewise, the total nitrogen in trout diets can replace over 50% with bakers' yeast *S. cerevisiae* nitrogen without changing the growth (Rumsey *et al.*, 1990). However, FM replacement with brewers' yeast at low levels in fish feeds showed best performances while no significant effect up to 45-50% replacement levels. In this study, 50% brewers' yeast replacement level significantly affected growth. Rumsey *et al.*(1990) mentioned that fish would not perform well, if brewers' yeast is the only protein source, because brewers' yeast is deficient in essential amino acids, vitamins, minerals, and nucleic acids. So high supplement levels of brewers' yeast in fish diets reduces growth and feed intake of fish (Gumus *et al.*, 2016; Ozório *et al.*, 2010).

The fish meal provides essential amino acids adequately in aquafeeds (NRC, 2011). Therefore, while partial replacement of fish meal by brewers' yeast increases growth performances, excessive levels could decrease their performance. For instance, alga Spirulina meal replaced 100% fish meal in sea cucumber, *Apostichopus japonicus* juvenile diets without affecting growth. Even though, sea cucumber juveniles showed

better growth performances at 25% fish meal replacement level with alga Spirulina meal (Tan *et al.*, 2009).

Conclusion

The optimum replacement percentage of CMFM[®] with brewers' yeast was 13.5% in juvenile *H. scabra* diets by the polynomial regression analysis of weight gain. Brewer's yeast could be a partial dietary replacer for high-value fishmeal in juvenile sea cucumber diets for maximizing growth and reducing production cost of sea cucumber feed in future.

Acknowledgements

The authors would like to thank Lion brewery (Ceylon) PLC, Colombo road, Biyagama, Sri Lanka for providing dry brewers' yeast.

References

Ajith Kumara, P.A.D., Jayanatha, J.S., Pushpakumara, J., Bandara, W. and Dissanayake, D.C. T. (2013). Artificial breeding and larval rearing of three tropical sea cucumber species – *Holothuria* scabra, *Pseudocolochirus violaceus* and *Colochirus quadrangularis* – in Sri Lanka. *SPC Beche- de-Mer Information Bulletin*, **33**: pp. 30-37.

Battaglene, S.C., Evizel Seymour, J. and Ramofafia, C. (1999). Survival and growth of cultured juvenile sea cucumbers, *Holothuria scabra*. *Aquaculture*, **178** (**3-4**): pp. 293-322. [Available at: <u>https://doi.org/10.1016/S0044-8486(99)00130-1</u>]

Bourdichon, F., Casaregola, S., Farrokh, C., Frisvad, J.C., Gerds, M.L., Hammes, W.P., Harnett, J., Huys, G., Laulund, S., Ouwehand, A., Powell, I.B., Prajapati, J.B., Seto, Y., Ter Schure, E., Van Boven, A., Vankerckhoven, V., Zgoda, A., Tuijtelaars S., Hansen E.B.(2012). Food fermentations: Microorganisms with technological beneficial use. *International Journal of Food Microbiology*, **154** (3): pp. 87-97.[Available at: https://doi.org/10.1016/j.ijfoodmicro.2011.12.030]

Chen, D. and Ainsworth, A. J. (1992). Glucan administration potentiates immune defence mechanisms of channel catfish, *Ictalurus punctatus* Rafinesque. *Journal of Fish Diseases*, **15** (4): pp. 295-304. [Available at: <u>https://doi.org/10.1111/j.1365-2761.1992.tb00667.x</u>]

Couttau, P. and Lavens, P. (1989). The use of yeast as single sell protein in aquacultural diets. *Mededelingen van de Faculteit Landbouwwetenschappen, Rijksuniversiteit Gent*, **54 (4b):** pp. 1583–1592.

(DOF) Department of Fisheries. (2019). Annual Fisheries Statistics. Retrieved November 14, 2020, from
 <u>https://www.fisheriesdept.gov.lk/web/images/pdf/downloads/FISHERIES_STATISTICS_</u>
 <u>2019_FINAL_PDF_compressed.pdf</u>

Essa, M. A., Mabrouk, H. A., Mohamed, R. A. and Michael, F. R. (2011). Evaluating different additive levels of yeast, *Saccharomyces cerevisiae*, on the growth and production performances of a hybrid of two populations of Egyptian African catfish, *Clarias gariepinus*. *Aquaculture*, **320** (1-2): pp. 137–14. [Available at: <u>https://doi.org/10.1016/j.aquaculture.2011.08.015</u>]

Fan, Y. J., Li, X. D., Luo, Z. and Song, Z. Z. (2010). Effects of replacement of dietary fish meal by soybean meal on growth, body composition and digestive enzyme activities in sea cucumber *Apostichopus japonicus* juveniles. *Journal of Dalian Fisheries University*, **25** (1): pp. 71-75.

Gumus, E., Aydin, B. and Kanyilmaz, M. (2016). Growth and feed utilization of goldfish (*Carassius auratus*) fed graded levels of brewers yeast (*Saccharomyces cerevisiae*). *Iranian Journal of Fisheries Sciences*, **15** (3): pp. 1124–1133.

Hamel, J. F., Conand, C., Pawson, D. L. and Mercier, A. (2001). The sea cucumber *Holothuria scabra* (Holothuroidea : Echinodermata): Its biology and exploitation as beche-de-mer. *Advances in Marine Biology*, **41**: pp. 129–223.

James, D.B. (2004). Captive breeding of the sea cucumber, *Holothuria scabra*, from India. In: Advances in Sea Cucumber Aquaculture and Management. FAO Fisheries Technical Paper 463 eds. Lovatelli, A., Conand, C., Purcell, S., Uthicke, S., Hamel, J. F. and Mercier, A. pp. 385-395. FAO, Rome. Kumara, A. and Dissanayake, C. (2017). Preliminary study on broodstock rearing, induced breeding and grow-out culture of the sea cucumber *Holothuria scabra* in Sri Lanka. *Aquaculture Research*, **48** (3): pp. 1058–1069. [Available at: <u>https://doi.org/10.1111/are.12948</u>]

Li, P. and Gatlin, D. M. (2003). Evaluation of brewers yeast (*Saccharomyces cerevisiae*) as a feed supplement for hybrid striped bass (*Morone chrysops* x *M. saxatilis*). *Aquaculture* **219**: pp. 681-692. [Available at: https://doi.org/10.1016/S0044-8486(02)00653-1]

Li, P. and Gatlin, D. M. (2004). Dietary brewers yeast and the prebiotic GrobioticTM AE influence growth performance, immune responses and resistance of hybrid striped bass (*Morone chrysops* x *M. saxatilis*) to *Streptococcus iniae* infection. *Aquaculture*, **231:** pp. 445-456. [Available at: https://doi.org/10.1016/j.aquaculture.2003.08.021]

Liao, M., Ren, T., He, L., Han, Y. and Jiang, Z. (2015). Optimum dietary proportion of soybean meal with fish meal, and its effects on growth, digestibility, and digestive enzyme activity of juvenile sea cucumber *Apostichopus japonicus*. *Fisheries Science*, **81** (5): pp. 915-922. [Available at: <u>https://doi.org/10.1007/s12562-015-0916-1]</u>

NRC (National Research Council). (2011). Nutrient Requirements of Fish and Shrimp. National Academy Press, Washington DC.

Oliva-Teles, A., and Gonçalves, P. (2001). Partial replacement of fishmeal by brewers yeast (*Saccaromyces cerevisae*) in diets for sea bass (*Dicentrarchus labrax*) juveniles. *Aquaculture*, **202** (**3-4**): pp. 269-278. [Available at: <u>https://doi.org/10.1016/S0044-8486(01)00777-3</u>]

Ozório, R. O. A., Turini, B. G. S., Môro, G. V., Oliveira, L. S. T., Portz, L. and Cyrino, J. E. P. (2010). Growth, nitrogen gain and indispensable amino acid retention of pacu (*Piaractus mesopotamicus*, Holmberg (1887) fed different brewers yeast (*Saccharomyces cerevisiae*) levels. *Aquaculture Nutrition*, **16** (3): pp. 276-283. [Available at: <u>https://doi.org/10.1111/j.1365-2095.2009.00662.x</u>]

Prasad, L., Nayak, B. B., Srivastava, P. P., Reddy, A. K. and Kohli, M. P. S. (2013). Use of brewer's yeast *Saccharomyces cerevisiae* as growth promoter in giant freshwater prawn (*Macrobrachium rosenbergii* de Man) post larvae. *Turkish Journal of Fisheries and Aquatic Sciences*, **13**: pp. 447-452. [Available at: <u>https://doi.org/10.4194/1303-2712-v13</u>]

Purcell, S. W., Hair, C. A. and Mills, D. J. (2012). Sea cucumber culture, farming and sea ranching in the tropics: progress, problems and opportunities. *Aquaculture*, **368–369:** pp. 68–81. [Available at: <u>https://doi.org/10.1016/j.aquaculture.2012.08.053]</u>

Rumsey, G. L., Hughes, S. G. and Kinsella, J. L. (1990). Use of Dietary Yeast *Saccharomyces cerevisiae* Nitrogen by Lake Trout. *Journal of the World Aquaculture Society*, **21** (3): pp. 205-209. [Available at: <u>https://doi.org/10.1111/j.1749-7345.1990.tb01024.x</u>]

Sakai, M. (1999). Current research status of fish immunostimulants. *Aquaculture*, **172:** pp. 63-92. [Available at: <u>https://doi.org/10.1016/S0044-8486(98)00436-0</u>]

Sanderson, G. W. and Jolly, S. O. (1994). The value of Phaffia yeast as a feed ingredient for salmonid fish. *Aquaculture*, **124** (**1-4**): pp. 193-200. [Available at: <u>https://doi.org/10.1016/0044-8486(94)90377-8]</u>

Siwicki, A. K., Anderson, D. P. and Rumsey, G. L. (1994). Dietary intake of immunostimulants by rainbow trout affects non-specific immunity and protection against furunculosis. *Veterinary Immunology and Immunopathology*, **41** (1-2): pp. 125-139. [Available at: https://doi.org/10.1016/0165-2427(94)90062-0]

Sun, Y., Chang, A. K., Wen, Z., Li, Y., Du, X. and Li, S. (2014). Effect of replacing dietary fish meal with silkworm (*Bombyx mori* L) caterpillar meal on growth and non-specific immunity of sea cucumber *Apostichopus japonicus* (Selenka). *Aquaculture Research*, **45:** pp. 1246-1252. [Available at: <u>https://doi.org/10.1111/are.12068]</u>

Tacon, A. G. J. (1995). Feed ingredients for carnivorous fish species: alternatives to fishmeal and other fishery resources. In: Sustainable Fsh Farming eds. Reinertsen, H. and Haaland, H. pp. 88-92. A.A. Balkema, Rotterdam.

Tan, X.Y., Luo, Z., Li, X. D., Zhang, S. L. and Sun, Z. Z. (2009). Effects of dietary fish meal replacement by different levels of alga Spirulina meal on growth performance and body composition of sea cucumber *Apostichopus japonicus*. *Journal of Dalian Fisheries University*, **24** (1): pp. 559-562.

Thanardkit, P., Khunrae, P., Suphantharika, M. and Verduyn, C. (2002). Glucan from spent brewer's yeast: preparation, analysis and use as a potential immunostimulant in shrimp feed. *World Journal of Microbiology and Biotechnology*, **18** (6): pp. 527-539. [Available at: <u>https://doi.org/10.1023/A:1016322227535</u>]

Tukmechi, A., Rahmati Andani, H. R., Manaffar, R. and Sheikhzadeh, N. (2011). Dietary administration of beta-mercapto-ethanol treated *Saccharomyces cerevisiae* enhanced the growth, innate immune response and disease resistance of the rainbow trout, *Oncorhynchus mykiss. Fish and Shellfish Immunology*, **30** (3): pp. 923-928. [Available at: https://doi.org/10.1016/j.fsi.2011.01.016]

Yang, G., Xu, Z., Tian, X., Dong, S. and Peng, M. (2015). Intestinal microbiota and immune related genes in sea cucumber (*Apostichopus japonicus*) response to dietary β-glucan supplementation. *Biochemical and Biophysical Research Communications*, **458** (1): pp. 98-103. [Available at: <u>https://doi.org/10.1016/j.bbrc.2015.01.074</u>]